

A new species and two new records of the genus *Alysia* Latreille (Hymenoptera, Braconidae, Alysiinae) from South Korea

Ju-Hyeong Sohn¹, Cornelis van Achterberg^{2,3}, Hyojoong Kim¹

1 Animal Systematics Lab., Department of Biological Science, Kunsan National University, Gunsan, 54150, Republic of Korea **2** State Key Laboratory of Rice Biology and Ministry of Agriculture, Zhejiang University, Hangzhou, 310058, China **3** Key Laboratory of Agricultural Entomology, Institute of Insect Science, Zhejiang University, Hangzhou, 310058, China

Corresponding author: Hyojoong Kim (hkim@kunsan.ac.kr)

Academic editor: J. Fernandez-Triana | Received 14 November 2022 | Accepted 13 January 2023 | Published 17 February 2023

<https://zoobank.org/CF9F06EC-165D-428F-9192-95F0C5FC2926>

Citation: Sohn J-H, van Achterberg C, Kim H (2023) A new species and two new records of the genus *Alysia* Latreille (Hymenoptera, Braconidae, Alysiinae) from South Korea. Journal of Hymenoptera Research 95: 45–58. <https://doi.org/10.3897/jhr.95.97527>

Abstract

In the genus *Alysia* Latreille, 1804 (Braconidae: Alysiinae), a new species, *Alysia erecta* **sp. nov.**, and two new records, *Alysia hebeiensis* Zhu, van Achterberg & Chen, 2018 and *A. sirin* Belokobylskij, 1998, are described and illustrated. In addition, the DNA barcode region of the mitochondrial *subunit I* (*COI*) of these species have been sequenced. An identification key for all *Alysia* species officially recorded from Korea is provided.

Keywords

Alysiini, COI, Hymenoptera, new combination, new record, new species, taxonomy

Introduction

The subfamily Alysiinae is a large taxon of the family Braconidae, consisting of over 2,440 valid species worldwide (Yu et al. 2016). Among them, 180 species in 21 genera are recorded in Korea (NIBR 2021). Alysiinae is generally distinguished from other subfamilies morphologically by having the exodont (= non-overlapping in closed condition) mandibles. Alysiinae includes two tribes, Alysiini and Dacnusiini, which are distinguishable each other in most cases by having vein r-m of fore wing present

Copyright Ju-Hyeong Sohn et al. This is an open access article distributed under the terms of the Creative Commons Attribution License (CC BY 4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

(Alysiini) or absent (Dacnusiini). Alysiinae belongs to the cyclostome clade, of which members are koinobiont endoparasitoids for dipterous larvae. They use the outward-curved teeth of the exodont mandibles to break out the host puparium (Docavo et al. 2002). Some species have been used for biological control (Ozawa et al. 2001; Chabert et al. 2012).

The genus *Alysia* Latreille, 1804, is a large group in the subfamily Alysiinae, including 125 species worldwide (Yu et al. 2016; Zhu et al. 2018). This genus can be diagnosed by having the first flagellomere longer than the second flagellomere (but not over 1.5 times), the comparatively short vein 3-SR of fore wing, the posterior position of vein r of fore wing, the propodeum more or less wrinkled or rugose and usually lacking an areola and vein m-cu of the hind wing distinct. According to Bartlett et al. (1978), *Alysia manducator* has been introduced to control the sheep blowfly, *Calliphora stygia* (Fabricius, 1781) in eastern Australia and New Zealand, but became established only in the latter country.

Comparatively, few papers are dealing with the eastern Palaearctic species: Two new species from Mongolia by Papp (1991); 14 new species from Far East Russia by Belokobylskij (1998); six new species from China by Zhu et al. (2018). Since seven species had been known in Korea, one species was recently transferred to the genus *Cratospila* (Sohn et al. 2022). Although Papp (1994) has been reported *A. brachycera*, *A. lucia*, *A. nigritarsis*, *A. sophia*, *A. tipulae* and *A. truncator* from Ryanggang, North Korea, the national checklist of South Korea (NIBR 2021) lists only two species (*A. sophia* and *A. tipulae*). Therefore, five species officially are recorded in South Korea from this study.

In this study, we present new morphological characters and the COI barcoding sequences of one new and two newly recorded species (*A. hebeiensis* and *A. sirin*). This study also provides descriptions, diagnosis, identification key and photographs of the diagnostic characters for the three species.

Materials and methods

Samples used in this study were collected with Malaise traps in South Korea at the Dodae-ri, buk-myeon, Gapyeong-gun, Gyeonggi-do. Sorting and preparation were done at the Animal Systematics Lab. (ASL), Department of Biological Science, Kunsan National University (KSNU). For morphological identification, Wharton et al. (1997) and Zhu et al. (2017) were used. Morphological characters were observed with a Leica M205C stereo microscope. The Taxapad database (Yu et al. 2016) was used for references. The terminology was followed of Wharton (2002) and van Achterberg (1993). The holotype of new species is deposited in the NIBR (National Institute of Biological Resources, Incheon) collection.

A Leica DMC2900 digital camera and a Leica M205 C microscope (Leica Geosystems AG, Mannheim, Germany) were used for photography; several pictures were taken for each final photo using multi-focusing technology. LAS V4.11 (Leica

Geosystems AG, St. Gallen, Switzerland) and HeliconFocus 7 (Helicon Soft, Kharkiv, Ukraine) software were used for stacking the photos. The final illustrations were created using Adobe Photoshop CS6.

Extraction of DNA was done in ASL, KSNU. Whole genomic DNA was extracted from the specimens by using a Labopass Tissue kit (COSMOgenetech, Daejeon, Korea) following the manufacturer's protocol. In order to conserve morphologically complete voucher specimens, DNA extraction method was used slightly modified from 'non-destructive method' by Favret (2005) and 'freezing method' by Yaakop et al. (2009). In the original protocol, the sample was crushed or wounded, and then soaked with 180 μ l of buffer ATL + 20 μ l of proteinase, following by three hours over incubation at 55 °C. In the slightly modified DNA extraction methods, samples were soaked with 180 μ l of buffer ATL + 20 μ l of proteinase K without destroying the sample, followed by 10 minutes incubation at 55 °C and then kept in a freezer at -22 °C overnight. After that the general protocol was used for the remaining steps. The primer set of LCO-1490 (5'-GGTCAACAATCATAAAGATATTGG-3') and HCO-2198 (5'-TAAACTTCAGGGTGACCAAAAAATCA-3') was used to amplify approximately 658 bp as the partial front region of the *COI*. The polymerase chain reaction (PCR) products were amplified by using AccuPowerH PCR PreMix (BIONEER, Corp., Daejeon, Korea) in 20 μ l reaction mixtures containing 0.4 μ M of each primer, 20 μ M of the dNTPs, 20 μ M of the MgCl₂, and 0.05 μ g of the genomic DNA template. PCR amplification was performed using a GS1 thermo-cycler (Gene Technologies, Ltd., Somerset, U.K) according to the following procedure: initial denaturation at 95 °C for 5 min, followed by 34 cycles at 94 °C for 35 sec; an annealing temperature of 48 °C for 25 sec; an extension at 72 °C for 45 sec, and a final extension at 72 °C for 5 min. The PCR products were visualized by electrophoresis on a 1.5% agarose gel. A single band was observed and then sequenced using an automated sequencer (ABI Prism 3730 XL DNA Analyzer, California, USA) at Macrogen Inc. (Seoul, South Korea).

Sequence alignment was performed in MEGA version 7 (Kumar et al. 2016) with ClustalW method. To estimate the pairwise genetic distances, the *P*-distance model was conducted using MEGA version 7.

Results and discussion

A total of 621 bp of the *COI* barcode region were sequenced for *Alysia erecta* sp. nov. (GenBank accession no. [OP391515](#)), *A. hebeiensis* Zhu & van Achterberg, 2018 (GenBank accession no. [OP391514](#)), and *A. sirin* Belokobylskij, 1998 (GenBank accession no. [OP391516](#)) Pairwise genetic distances were calculated by using 'P-distance' model with option for pairwise deletion; *A. erecta* differed by 6% from *A. hebeiensis* and by 9% from *A. sirin*; *A. hebeiensis* 10% from *A. sirin*.

Taxonomy

Alysia Latreille, 1804

Figs 1–3

Alysia Latreille, 1804: 173–174; Shenefelt 1974: 939; Wharton 1980: 458; Chen and Wu 1994: 28; Belokobylskij 1998: 170; Zhu et al. 2018: 2. Type species: *Ichneumon manducator* Panzer, 1799.

Cechenus Illiger, 1807: 54; Type species: *Ichneumon manducator* Panzer, 1799. Synonymized by Curtis 1826.

Bassus Nees, 1812: 201; Type species: *Ichneumon manducator* Panzer, 1799. Synonymized by Nees 1819.

Anarcha Foerster, 1863: 265; Ashmead 1900: 105; Baltazar 1962: 759. Type species: *Anaraha notabilis* Foerster, 1863. Synonymized by Fischer 1971.

Goniarcha Foerster, 1863: 265; Marshall 1872: 125; Ashmead 1900: 105. Type species: *Alysia lucicola* Haliday, 1838. Synonymized by Marshall 1894.

Strophaea Foerster, 1863: 265; Marshall 1872: 127; Ashmead 1900: 105. Type species: *Alysia rufidens* Nees, 1834. Synonymized by Marshall 1894.

Diagnosis. First flagellomere longer than second (Figs 1B, J, 2B, J, 3B, J), not over 1.6 times, face granulate (Figs 1E, 2E, 3E) or largely smooth, eye slightly oval, clypeus triangularly shaped, wide and protruding anteriorly (Figs 1E, 2E, 3E); mandible (Figs 1K, L, 2K, L, 3K, L) with 3 teeth, third mostly lobe-shaped. second tooth narrow and sharp; pronope absent, notauli present, precoxal sulcus distinct, medially deeply impressed (Figs 1G, 2G, 3G), scutellar sulcus distinct, propodeum more or less rugose and usually without areola, sometimes with enlarged spiracles; pterostigma robust, fore wing (Figs 1C, 2C, 3C) vein 2-SR slightly bent, first discal cell shorter than wide in median length. vein 3-SR usually shorter than vein 2-SR; veins 2-SR+M and r-m not sclerotized, hind wing vein 1r-m shorter than vein M+CU, vein m-cu distinct; first tergite with dorsope (Figs 1H, 2H, 3H).

Biology. Endoparasitoids of larval Calliphoridae, Sarcophagidae, Tephritidae, Anthomyiidae, Agromyzidae and Mycetophylidae (Yu et al. 2016).

Distribution. Cosmopolitan, but most *Alysia* species occur in the northern part of the Northern Hemisphere and many are Holarctic. About 70% of the species have most or all of their range within the boreal coniferous biome (Wharton 1986).

Key to species of *Alysia* Latreille from South Korea

- 1 Antenna with 5–10 white segments subapically (Fig. 3B); first metasomal tergite 2.4–3.0 times longer than its apical width (Fig. 3H) *A. sirin* Belokobylskij, 1998
- Antenna without white segments (Fig. 2B); first tergite 1.0–1.7 times longer than its apical width (Fig. 2H) 2

- 2 First flagellomere about 1.5 times longer than second; setose part of ovipositor sheath 1.6–1.7 times longer than hind tibia; eye in dorsal view 1.1–1.2 times as long as temple.....*A. tipulae* (Scopoli, 1763)
- First flagellomere 1.2–1.3 times longer than second; setose part of ovipositor sheath 0.5–1.3 times as long as hind tibia; eye in dorsal view 1.2–1.4 times as long as temple..... **3**
- 3 Setose part of ovipositor sheath 1.2 times longer than hind tibia (Fig. 2A); hind femur 4.4–4.6 times longer than its maximum width.....
..... *A. hebeiensis* Zhu, van Achterberg & Chen, 2018
- Setose part of ovipositor sheath 0.5–0.7 times as long as hind tibia; hind femur 3.9–4.0 times longer than its maximum width..... **4**
- 4 First antennal flagellomere about 2.5 times longer than wide (Fig. 2J); mandible 1.2 times longer than its maximum width (Fig. 1K); pterostigma dark brown; metasoma after first tergite dark brown (Fig. 1A)*A. erecta* sp. nov.
- First flagellomere about 3.5 times longer than wide; mandible 1.6–1.7 times longer than its maximum width; pterostigma pale yellowish brown to brown; metasoma after first tergite usually yellow or orange.....
.....*A. sophia* (Haliday, 1838)

***Alysia erecta* Sohn & van Achterberg, sp. nov.**

<https://zoobank.org/78EBA32D-8A2D-4D4E-8C95-904A7289EA87>

Fig. 1A–L

Type material. *Holotype*, ♀ (NIBR), **South Korea**, Dodae-ri, buk-myeon, Gapyeong-gun, Gyeonggi-do, 37°56'11.8"N, 127°28'50.2"E, 05.IV.2018, Sohn. GenBank accession no. [OP391515](https://doi.org/10.21203/rs.3.rs-151515).

Comparative diagnosis. The new species is recognizable by its comparatively short ovipositor sheath (setose part 0.6 times longer than mesosoma *versus* 1.0–1.7 times in other S Korean species), the short first flagellomere (2.5 times longer than wide *versus* 3.0–4.5 times) and robust mandible (1.2 times longer than wide *versus* 1.4–1.7 times). The new species runs in the key by Zhu et al. (2018) to *A. hebeiensis* Zhu & van Achterberg, 2018 and differs from this species by having the first flagellomere less slender (2.5 times *versus* 3.8–4.0 times in *A. hebeiensis*) and the first metasomal tergite shorter (1.1 times longer than its apical length *versus* 1.2–1.7 times).

The new species runs in the key by Belokobylskij (1998) to *A. masneri* Wharton, 1988 based on the colour of the clypeus, but to *A. vladik* Belokobylskij, 1998 if only morphological characters are used. The new species differs from *A. masneri* by the robust first tergite (1.1 times longer than its apical width *versus* 1.6 times in *A. masneri*), the shorter antenna (0.9 times as long as body *versus* distinctly longer than body) and the shorter ovipositor sheath (0.6 times as long as mesosoma *versus* 0.9 times). The new species differs from *A. vladik* by having the first flagellomere less slender (2.5 times *versus* 4.0 times in *A. vladik*), the hind femur less slender (3.9 times *versus* 4.3 times), the first

metasomal tergite shorter (1.1 times longer than its apical width *versus* 1.2–1.3 times), the clypeus black (similar to colour of face *versus* yellowish, contrasting with black face).

According to the key by Wharton (1986) the new species belongs to his *Alysia tipulae* group of the subgenus *Anarcha* Foerster. It runs in the key by Wharton (1988) to *A. umbrata* Stelfox, 1941 and differs by having the mandible less slender (1.2 times *versus* 1.6–1.7 times in *A. umbrata*).

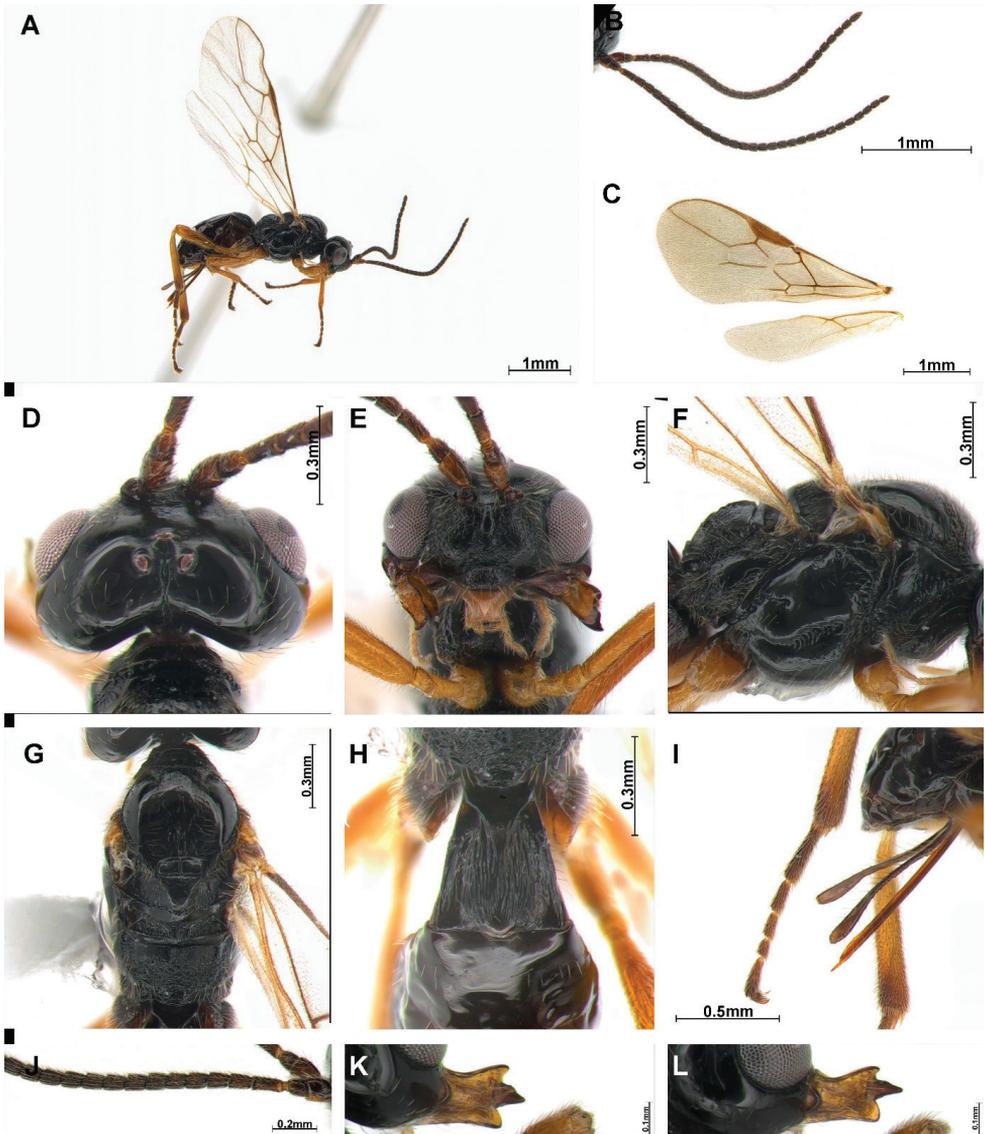


Figure 1. A–L *Alysia erecta* sp. nov., ♀ **A** habitus, lateral view **B** antennae **C** wings **D** head, dorsal view **E** head, front view **F** mesosoma, dorsal view **G** mesosoma, lateral view **H** anterior half of metasoma, dorsal view **I** ovipositor sheath, lateral view **J** basal part of antenna **K, L** mandible.

Description. Holotype, ♀, length of body 3.1 mm in lateral view, length of antenna 3.0 mm and of fore wing 3.6 mm.

Colour: Body (Fig. 1A) black, first tergite and mesonotum entirely reddish brown; antenna dark brown; mandible pale yellow; leg yellowish brown basally, tarsus brown.

Head (Fig. 1D): Width of head 1.9 times its median length in dorsal view. Antenna 0.9 times longer than body, 31 segmented. First flagellomere 1.3 times longer than second and 2.5 times longer than wide. Compounded eye slightly oval, in lateral view 1.6 times as long as wide. Minimum width of face (Fig. 1E) 1.6 times its height (measured from ventral rim of antennal sockets to upper margin of clypeus); face setose, wrinkled and rather mat. Eye in dorsal view 1.4 times as long as temple. Frons nearly entirely glabrous. Ocello-ocular line (OOL) 4.1 times longer than diameter of anterior ocellus; OOL: antero-posterior ocellar line (AOL): postero-ocellar line (POL) = 24 : 7 : 10. Stemmaticum with deep and long median groove. Vertex smooth and with sparse setae. Mandible 1.2 times longer than wide, with three teeth; first tooth curved, with distinct incision between first and second tooth; second tooth reddish brown, narrow and sculptured; second tooth 1.5 times longer than first tooth. Maxillary palp pale yellow and 0.4 times longer than mesosoma.

Mesosoma: In dorsal view mesosoma 1.9 times longer than wide, 1.4 times longer than wide in lateral view. Mesoscutum (Fig. 1G) with oval medio-posterior depression and with setae; notauli impressed anteriorly, not reaching medio-posterior depression; anterior mesosoma crenulated; scutellar sulcus with two carinae; in lateral view, apical part of mesopleuron and metapleuron with long setae. metanotum sculptured. Propodeum (Fig. 1G) entirely wrinkled, 0.4 times longer than wide in maximum length; precoxal sulcus (Fig. 1F) deep and distinct, occupying entire length of mesopleuron; propodeum curved dorsally in lateral view. Fore wing (Fig. 1C) 2.3 times as long as wide in maximum length; pterostigma 3.9 times as long as wide; vein r of fore wing 2.6 times longer than wide and issued from distal third of pterostigma; vein 2-SR slightly bent; vein 3-SR 1.1 times longer than vein 2-SR; vein 2-SR+M and r-m not sclerotized; 2-SR: r : 3-SR = 28: 5: 25; first discal cell of fore wing approx. 0.8 times longer than wide in median length; first subdiscal cell of fore wing approx. 3.4 times as long as wide in median length. Hind wing vein M+CU: vein 1r-m = 9: 7.

Leg: Hind coxa reddish brown apically, compressed and 1.1 times longer than hind trochanter; hind femur brownish yellow, 3.9 times as long as wide and 0.7 times longer than hind tibia; hind tibia as long as hind tarsus.

Metasoma: First tergite (Fig. 1H) striate, 1.1 times longer than its apical width and blackish. Setose part of ovipositor sheath (Fig. 1I) 0.6 times as long as mesosoma and 0.5 times as long as hind tibia. Ovipositor without subapical dorsal notch (Fig. 1I).

Male. Unknown.

Distribution. South Korea.

Etymology. Named after the erect setae on the flagellomeres: “*erecta*” is Latin for erect.

***Alysia hebeiensis* Zhu, van Achterberg & Chen, 2018**

Fig. 2A–L

Alysia hebeiensis Zhu, van Achterberg & Chen, 2018: 4

Material. 1♀ (NIBR), **South Korea**, Dodae-ri, buk-myeon, Gapyeong-gun, Gyeonggi-do, 37°56'11.8"N, 127°28'50.2"E, 04.IV.2018, Sohn. GenBank accession no. [OP391514](#).

Redescription. ♀, length of body in lateral view 3.4 mm, length of antenna 4.0 mm and of fore wing 4.1 mm.

Colour: Body (Fig. 2A) black, but metasoma entirely reddish brown; antenna dark brown basally, leg yellowish brown basally, tarsus brown.

Head (Fig. 2D): Width of head 2.0 times its median length in dorsal view. Antenna 1.2 times longer than body, 38 segmented. First flagellomere 1.2 times longer than second and 3.8 times longer than wide. Compounded eye slightly oval, in lateral view 1.4 times as long as wide. Minimum width of face (Fig. 2E) 1.8 times its height (measured from ventral rim of antennal sockets to upper margin of clypeus); face wrinkled with long setae. Eye in dorsal view 1.3 times as long as temple. Ocello-ocular line (OOL) 6.1 times longer than diameter of anterior ocellus; OOL: antero-posterior ocellar line (AOL) : postero-ocellar line (POL) = 23 : 6 : 7. Vertex smooth and with sparse long setae. Mandible 1.4 times longer than wide, with three teeth; first tooth lobe-shaped; second tooth reddish brown, wide and sculptured; second tooth 1.3 times longer than first tooth. Maxillary palp pale yellow and 0.5 times longer than mesosoma.

Mesosoma: In dorsal view mesosoma 2.0 times longer than wide, 1.6 times longer than wide in lateral view. Mesoscutum (Fig. 2G) with oval medio-posterior depression and long setae; notauli impressed anteriorly, not reaching medio-posterior depression; anteriorly mesosoma crenulated widely; scutellar sulcus with four carinae; in lateral view, mesopleuron and metapleuron with long setae. Metanotum sculptured. Propodeum (Fig. 2G) entirely wrinkled, 0.5 times longer than wide in maximum length; precoxal sulcus (Fig. 2F) distinct, without setae, occupying entire length of mesopleuron; propodeum curved dorsally in lateral view. Fore wing (Fig. 2C) 2.2 times as long as wide in maximum length; pterostigma 3.4 times as long as wide; vein r of fore wing 2.1 times longer than wide; vein 2-SR slightly bent; vein 2-SR+M and r-m not sclerotized; 2-SR: r : 3-SR = 6 : 1 : 5; first discal cell of fore wing approx. 0.8 times longer than wide in median length; first subdiscal cell of fore wing approx. 3.6 times as long as wide in median length. Hind wing vein M+CU: vein 1r-m = 4: 1.

Leg: Hind coxa apically pale yellow and 1.2 times longer than hind trochanter; hind femur 4.4 times as long as wide and 0.7 times longer than hind tibia; hind tibia 1.2 times longer than hind tarsus.

Metasoma: First tergite striate and narrow, 1.7 times longer than its apical width and dark brown. Setose part of ovipositor sheath (Fig. 2I) 1.2 times longer than mesosoma and 1.2 times longer than hind tibia.

Male. Unknown.

Distribution. China (Zhu et al. 2018), South Korea (new record).

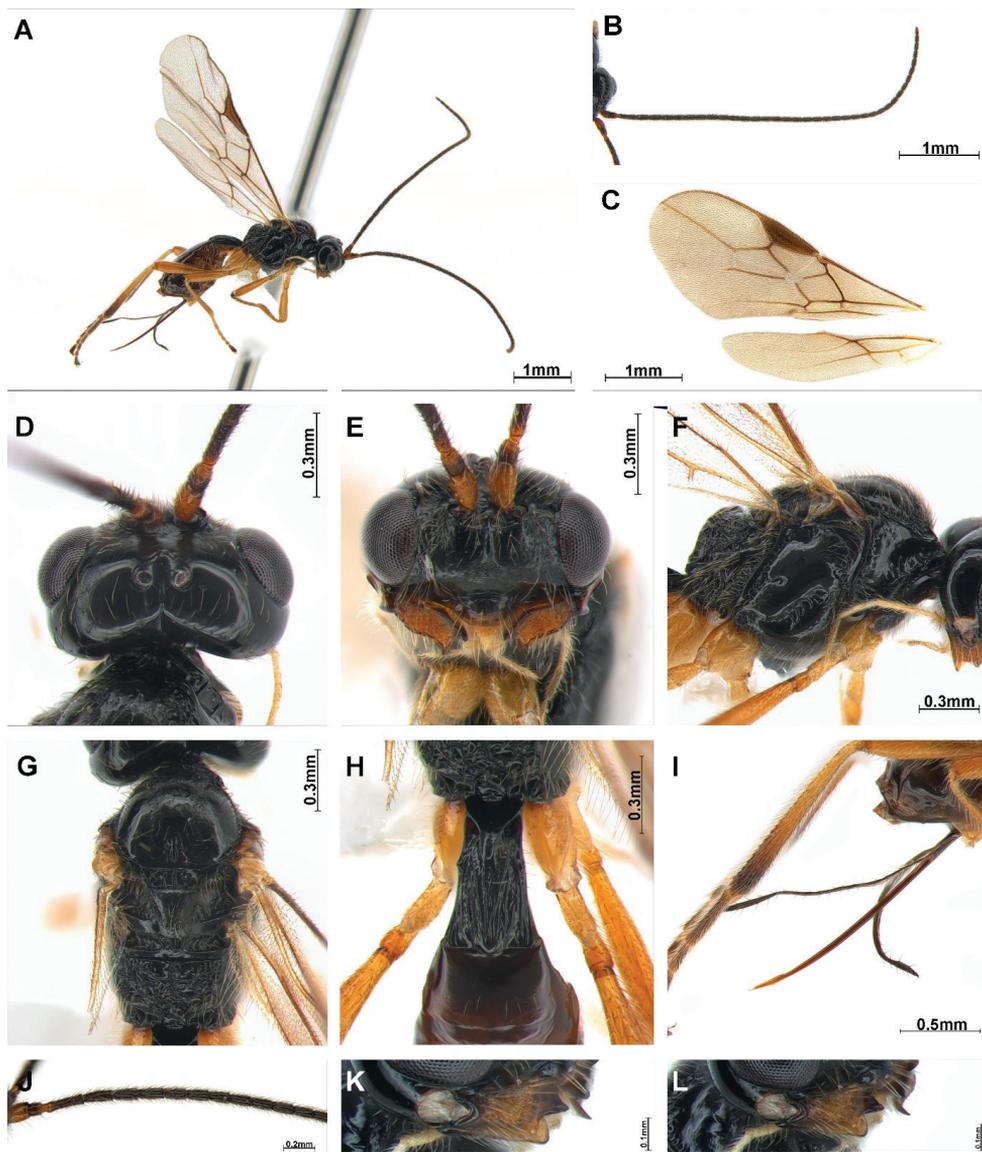


Figure 2. A–L *Alysia hebeiensis* Zhu, van Achterberg & Chen, 2018 ♀ **A** habitus, lateral view **B** antennae **C** wings **D** head, dorsal view **E** head, front view **F** mesosoma, dorsal view **G** mesosoma, lateral view **H** anterior half of metasoma, dorsal view **I** ovipositor sheath, lateral view **J** basal part of antenna **K**, **L** mandible.

Table 1. COI pairwise genetic distances between the three *Alysia* spp. from South Korea.

	<i>A. erecta</i>	<i>A. hebeiensis</i>	<i>A. sirin</i>
<i>A. erecta</i>	0.000		
<i>A. hebeiensis</i>	0.061	0.000	
<i>A. sirin</i>	0.094	0.098	0.000

***Alysia sirin* Belokobylskij, 1998**

Fig. 3A–L

Alysia sirin Belokobylskij, 1998: 178.

Material. 1♀ (NIBR), **South Korea**, Dodae-ri, buk-myeon, Gapyeong-gun, Gyeonggi-do, 37°56'11.8"N, 127°28'50.2"E, 05.IV.2018, Sohn. GenBank accession no. [OP391516](#).

Description. ♀, length of body in lateral view 3.9 mm, length of antenna 4.0 mm and of fore wing 3.5 mm.

Colour: Body (Fig. 3A) black, but metasoma entirely pale yellow; antenna dark brown basally, apical parts pale yellow (two apical segments missing); hind leg basally tri-coloured, coxa pale yellow, apical part of hind femur and hind tibia yellowish brown, posterior part of hind femur, hind tibia and hind tarsus reddish brown.

Head (Fig. 3D): Width of head 2.1 times its median length in dorsal view. Antenna incomplete, remaining part as long as body and 29 segmented. First flagellomere 1.4 times longer than second and 4.4 times longer than wide. Compounded eye slightly oval, in lateral view 1.2 times as long as wide. Minimum width of face (Fig. 3E) 1.8 times its height (measured from ventral rim of antennal sockets to upper margin of clypeus); face granulate and with long setae; labrum wrinkled. Eye in dorsal view 2.4 times as long as temple. Ocello-ocular line (OOL) 5.4 times longer than diameter of anterior ocellus; OOL: antero-posterior ocellar line (AOL): postero-ocular line (POL) = 22: 5: 7. Vertex with long setae. Mandible 1.6 times longer than wide, first tooth with setae; first tooth lobe-shaped; second tooth reddish brown, narrow and sharp; second tooth 1.5 times longer than first tooth; apical part of third tooth reddish brown, short and flat. Maxillary palp pale yellow and 0.8 times longer than mesosoma.

Mesosoma: In dorsal view mesosoma 1.9 times longer than wide, 1.4 times longer than wide in lateral view. Mesoscutum (Fig. 3G) with slightly oval medio-posterior depression and long setae; notauli impressed anteriorly, not reaching medio-posterior depression; mesosoma crenulated anteriorly; scutellar sulcus with four carinae; in lateral view, apical part of mesopleuron and metapleuron with long setae. Metanotum rugose. Propodeum (Fig. 3G) entirely rugose, 0.4 times longer than wide in maximum length; precoxal sulcus (Fig. 3F) distinct, apical part with setae, occupying entire length of mesopleuron; propodeum curved dorsally in lateral view. Fore wing (Fig. 3C) 2.2 times as long as wide in maximum length; pterostigma 3.3 times as long as wide; vein r of fore wing 3.6 times longer than wide; vein 2-SR slightly bent; vein 2-SR+M and r-m not sclerotized; 2-SR: r : 3-SR = 10: 2: 7; first discal cell of fore wing approx. 0.9 times longer than wide in median length; first subdiscal cell of fore wing approx. 4.1 times longer than wide medially. Hind wing vein M+CU: vein 1r-m = 16: 5.

Leg: Hind coxa apically pale yellow; hind coxa 1.2 times longer than hind trochanter; hind femur 4.6 times as long as wide and 0.7 times longer than hind tibia; hind tibia as long as hind tarsus.

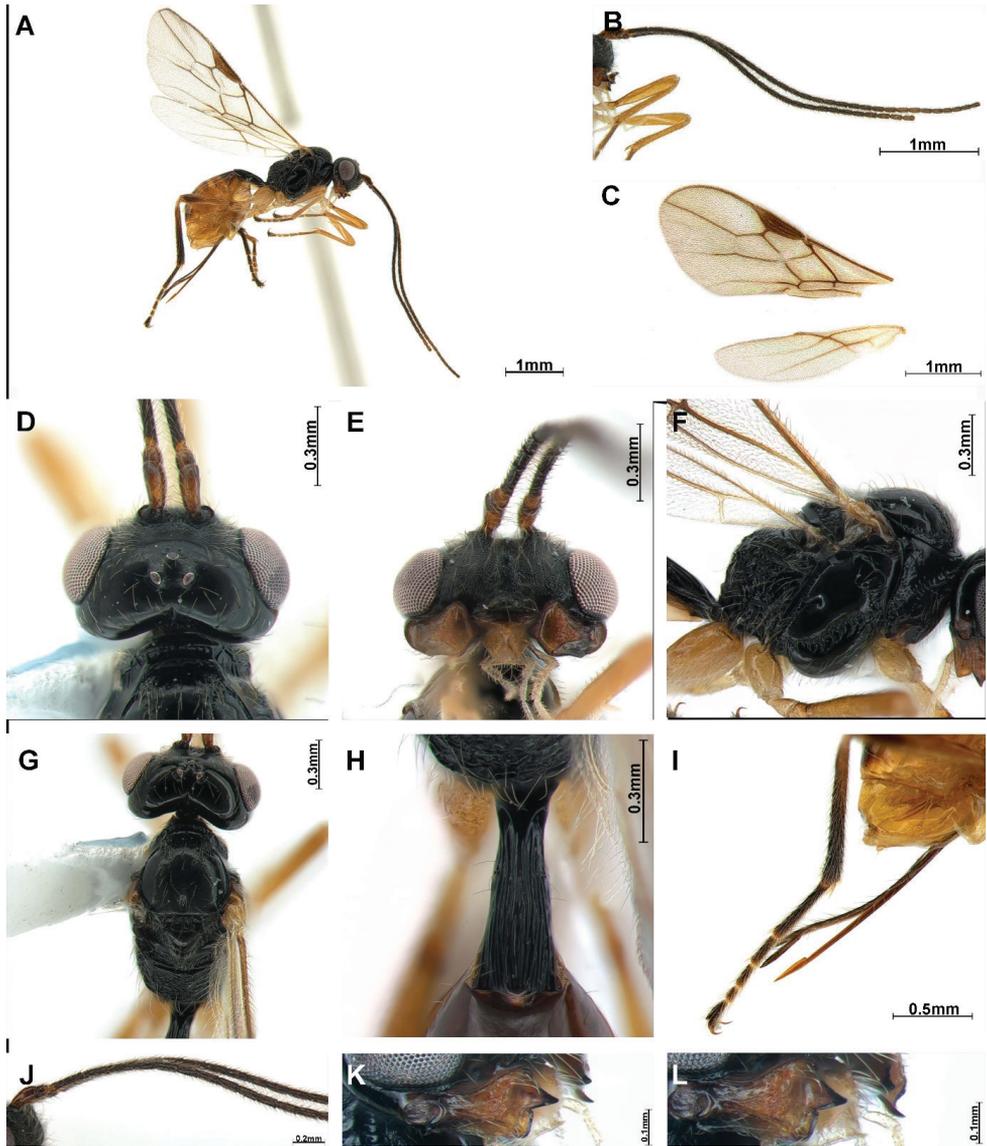


Figure 3. A–L *Alysia sirin* Belokobylskij, 1998♀ **A** habitus, lateral view **B** antennae **C** wings **D** head, dorsal view **E** head, front view **F** mesosoma, dorsal view **G** mesosoma, lateral view **H** anterior half of metasoma, dorsal view **I** ovipositor sheath, lateral view **J** basal part of antenna **K, L** mandible.

Metasoma: First tergite striate and narrow, 2.5 times longer than its apical width and dark brown. Setose part of ovipositor sheath (Fig. 3I) 1.3 times longer than mesosoma and 1.3 times longer than hind tibia.

Male. Unknown.

Distribution. Eastern Palearctic, Japan, Russia (Yu et al. 2016), South Korea (new record).

Acknowledgements

This work was supported by a grant from the National Institute of Biological Resources (NIBR), funded by the Ministry of Environment (MOE) of the Republic of Korea (NIBR202203201). It was also supported by Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Education (2022R1A2C1091308). It was also supported by a grant from the Korea Environment Industry & Technology Institute (KEITI) through Exotic Invasive Species Management Program (2018002270005) funded by Korea Ministry of Environment (MOE) of the Republic of Korea.

References

- Ashmead WH (1900) Classification of the Ichneumon flies, or the superfamily Ichneumonoidea. Proceedings of the United States National Museum. 23 (1206): 1–220. <https://doi.org/10.5479/si.00963801.23-1206.1>
- Baltazar CR (1962) The genera of parasitic Hymenoptera in the Philippines, Part 1. Pacific Insects 4(4): 737–771.
- Bartlett BR, Clausen CP, Debach P, Goerden RD, Legner EF, Mcmurtry JA, Oatman ER (1978) Introduced parasites and predators of arthropod pests and weeds: A world review. Agricultural Research Service. United States Department of Agriculture. Agriculture Handbook 480: 545.
- Belokobylskij SA (1998) Tribe Alysiini. In: Ler PA (Ed.) Key to the Insects of Russian Far East. Dal'nauka, Vladivostok, 163–298. [In Russian]
- Chabert S, Allemand R, Poyet M, Eslin P, Gibert P (2012) Ability of European parasitoids (Hymenoptera) to control a new invasive Asiatic pest, *Drosophila suzukii*. Biological Control 63: 40–47. <http://doi.org/10.1016/j.biocontrol.2012.05.005>
- Chen JH, Wu ZS (1994) The Alysiini of China: (Hymenoptera: Braconidae: Alysiinae). China Agricultural Press. Fuzhou, 218 pp. [in Chinese with English summary]
- Curtis J (1826) British Entomology; being illustrations and descriptions of the genera of insects found in Great Britain and Ireland. London 3: 120–141.
- Docavo I, Tormos J, Fischer M (2002) Three new species of *Chorebus* from Spain (Hymenoptera: Braconidae: Alysiinae). Florida Entomologist 85(1): 208–215. [https://doi.org/10.1653/0015-4040\(2002\)085\[0208:TNSOCF\]2.0.CO;2](https://doi.org/10.1653/0015-4040(2002)085[0208:TNSOCF]2.0.CO;2)
- Favret C (2005) New non-destructive DNA extraction and specimen clearing technique for aphids (Hemiptera). Proceedings of the Entomological Society of Washington 107: 469–470.
- Fischer M (1971) Untersuchungen über die europäischen Alysiini mit besonderer Berücksichtigung der Fauna Niederösterreichs (Hymenoptera, Braconidae). Polskie Pismo Entomologiczne 41: 19–160.
- Foerster A (1863) Synopsis der Familien und Gattungen der Braconiden. Verhandlungen des Naturhistorischen Vereins der Preussischen Rheinlande und Westfalens 19: 225–288.
- Haliday AH (1838) Essay on parasitic Hymenoptera. Entomological Magazine 5(3): 209–249.

- Illiger K (1807) IV. Vergleichung der Gattungen der Hautflügler Piezata Fabr. Hymenoptera Linn. Jur. Magazin für Insektenkunde 6: 189–199.
- Kumar S, Stecher G, Tamura K (2016) MEGA7: Molecular Evolutionary Genetics Analysis version 7.0 for bigger datasets. *Molecular Biology and Evolution* 33: 1870–1874. <https://doi.org/10.1093/molbev/msw054>
- Latreille PA (1804) Histoire naturelle, générale et particuliere, des Crustacés et des Insectes. Tome treizième. F. Dufart, Paris, 161–190.
- Marshall TA (1872) A catalogue of British Hymenoptera; Chrysididae, Ichneumonidae, Braconidae, and Evanidae. A. Napier. London. The Entomological Society of London, 136 pp.
- Marshall TA (1894) A monograph of the British Braconidae Part V. Transactions of the Royal Entomological Society of London 1894: 497–534. <https://doi.org/10.1111/j.1365-2311.1894.tb02098.x>
- Nees von Esenbeck CG (1812) Ichneumonides Adsciti, in Genera et Familias Divisi. *Magazin Gesellschaft Naturforschender Freunde zu Berlin* 6(1812): 183–221.
- Nees von Esenbeck CG (1819) Appendix ad J.L.C. Gravenhorst conspectum generum et familiarum Ichneumonidum, genera et familias Ichneumonidum adscitorum exhibens Isis. *Nova Acta Academia Leopoldina in Hedicke* 9(1818): 299–310.
- Nees von Esenbeck CG (1834) Hymenopterorum Ichneumonibus affinum, monographiae, genera Europaea et species illustrantes 2: 448. <https://doi.org/10.5962/bhl.title.26555>
- NIBR (2021) National List of Species of Korea. National Institute of Biological Resources, Incheon, 1–988. <http://kbr.go.kr> [accessed on 16.iii.2022]
- Ozawa A, Saito T, Ota M (2001) Biological control of the American Serpentine leafminer, *Liriomyza trifolii* (Burgess), on tomato in greenhouses by parasitoids. II. Evaluation of biological control by *Diglyphus isaea* (Walker) and *Dacnusa sibirica* Telenga in commercial greenhouses. *Japanese Journal of Applied Entomology and Zoology* 45(2): 61–74. <https://doi.org/10.1303/jjaez.2001.61>
- Panzer GWF (1799) Faunae Insectorum Germanicae. Heft 72. Felsecker, Nürnberg, 4 pp.
- Papp J (1991) Braconidae (Hymenoptera) from Mongolia, XI. *Acta Zoologica Hungarica* 37: 217–224.
- Papp J (1994) Braconidae (Hymenoptera) from Korea XV. *Acta Zoologica Academiae Scientiarum Hungaricae* 40(2): 133–156.
- Shenefelt RD (1974) Braconidae 7. Alysiinae. *Hymenopterorum Catalogus. Pars 11*: 985–1113.
- Sohn JH, van Achterberg C, Han YJ, Kim H (2021) Four new species of the genus *Cratospila* Foerster (Hymenoptera: Braconidae: Alysiinae) from South Korea. *Zookeys* 1022: 51–64. <https://doi.org/10.3897/zookeys.1022.62562>
- van Achterberg C (1993) Illustrated key to the subfamilies of the Braconidae (Hymenoptera: Ichneumonoidea). *Zoologische Verhandelingen Leiden* 283: 1–189.
- Wharton RA (1980) Review of the Nearctic Alysiini (Hymenoptera, Braconidae): with discussion of generic relationships within the tribe. University of California Press, Berkeley, USA 88: 1–112.
- Wharton RA (1986) The braconid genus *Alysia* (Hymenoptera): a description of the subgenera and a revision of the subgenus *Alysia*. *Systematic Entomology* 11(4): 453–504. <https://doi.org/10.1111/j.1365-3113.1986.tb00538.x>

- Wharton RA (1988) The braconid genus *Alysia* (Hymenoptera): a revision of the subgenus *Anarcha*. Contributions of the American Entomological Institute, USA 11(4): 453–504. <https://doi.org/10.1111/j.1365-3113.1986.tb00538.x>
- Wharton RA, Marsh P, Sharkey M (1997) Manual of the New World Genera of the Family Braconidae (Hymenoptera). The International Society of Hymenopterists. Washington D.C., 439 pp.
- Wharton RA (2002) Revision of the Australian Alysiini (Hymenoptera: Braconidae). Invertebrate Systematics 16: 7–105. <https://doi.org/10.1071/IT01012>
- Yaakop S, van Achterberg C, Idris A-B (2009) Heratemis Walker (Hymenoptera: Braconidae: Alysiinae: Alysiini): revision and reconstruction of the phylogeny combining molecular data and morphology. Tijdschrift voor Entomologie 152(1): 1–64. <https://doi.org/10.1163/22119434-900000268>
- Yu DSK, van Achterberg C, Horstmann K (2016) Taxapad 2016, Ichneumonoidea 2015. Database on flash-drive. Nepean, Ontario, Canada.
- Zhu J-C, van Achterberg C, Chen X-X (2017) An illustrated key to the genera and subgenera of the Alysiini (Hymenoptera, Braconidae, Alysiinae), with three genera new for China. ZooKeys 722: 37–79. <https://doi.org/10.3897/zookeys.722.14799>
- Zhu J-C, van Achterberg C, Chen X-X (2018) Review of the genus *Alysia* Latreille (Hymenoptera, Braconidae, Alysiinae), with description of six new species from China. Zootaxa 4500(1): 1–42. <https://doi.org/10.11646/zootaxa.4500.1.1>