# Sexual dimorphism in excess power index of four North American native bees (Hymenoptera, Andrenidae, Apidae, and Halictidae) 

Patrick A. Vigueira ${ }^{1^{*}}$, Cynthia C. Vigueira ${ }^{2}$, Joshua W. Campbell ${ }^{3^{*}}$, Samia Ladner ${ }^{4}$, Gabrielle Hayes ${ }^{4}$, Elizabeth Riser ${ }^{4}$<br>I Independent Researcher, Crestwood, MO, 63126, USA 2 Division of Biology and Biomedical Sciences, Washington University in St. Louis, St. Louis, MO 63130, USA 3 USDA-ARS Pest Management Research Unit, Northern Plains Agricultural Research Laboratory, Sidney, MT, 59270, USA 4 Department of Biology, High Point University, High Point, NC 27268, USA

Corresponding author: Joshua W. Campbell (Joshua.Campbell@usda.gov)

Academic editor: Jack Neff | Received 10 December 2022 | Accepted 31 March 2023 | Published 11 April 2023
https://zoobank.org/CCF95B60-A562-45E1-8B25-2F7233EAE096
Citation: Vigueira PA, Vigueira CC, Campbell JW, Ladner S, Hayes G, Riser E (2023) Sexual dimorphism in excess power index of four North American native bees (Hymenoptera, Andrenidae, Apidae, and Halictidae). Journal of Hymenoptera Research 96: 121-128. https://doi.org/10.3897/jhr.96.98652


#### Abstract

A multitude of hymenopteran species exhibit sexual dimorphism for simple traits, such as color, size, or antennal segment number. These differences can reflect selection for specialized biological roles, many of which have not been documented for the majority of bee and wasp species. The excess power index (EPI) is an estimate of insect flight performance that is inferred by the combination of several morphological characteristics. We compared the female and male EPIs in four species of native bees: Agapostemon virescens Fabricius, Andrena carlini Cockerell, Melissodes bimaculata Lepeletier and Xylocopa virginica L.. While females of each species had a significantly larger whole body mass, males had a significantly larger EPI. A larger body mass for females is associated with egg laying abilities and foraging behavior. Male fitness may be dependent on EPI; males that have greater flight capacity can travel further or remain in flight for longer time periods in search of mates.


## Keywords

excess power index, flight machinery, sexual dimorphism, wing loading

[^0]
## Introduction

In North America, it is estimated that there are 4,000 different species of native bees (Michener 2007). Native bees are important pollinators for a multitude of crops (AllenPerkins et al. 2022) and non-crop plants alike (Ollerton et al. 2011). However, native bees have been in decline due to numerous anthropogenic activities (Cameron et al. 2011; Abbate et al. 2019). Despite their economic and ecological importance, the basic biology and life history of the vast majority of native bee species have remained unstudied. Observational field studies of bees can be challenging, time consuming, and often unsuccessful at finding species of interest. One indirect method of learning about native bee biology is through the use of preserved specimens. Museum specimens have been used to assess population declines of native bees (Colla et al. 2012), pollen sources utilized (Kleijn and Raemakers 2008; Scheper et al. 2014), phylogenetics (Vaudo et al. 2018), and species determination via morphometrics and DNA barcoding (Ndungu et al. 2017).

Previous studies have used flight-relevant morphological features of bees to predict an individual's flight performance potential that is described in a composite value called Excess Power Index (EPI). Excess Power Index has been used to calculate the maximum power that an individual bee has in order to maintain a steady flight at equilibrium in relation to the bee's body proportions and wing dimensions (Hepburn et al. 1998). These measurements have been used to compare flight mechanics of workers from different honey bee subspecies (Apis mellifera subsp. L.), honey bee species (Apis spp. L.), and castes (Hepburn et al. 1998; Hepburn et al. 1999; Radloff et al. 2003). Additionally, EPI has been used to examine relationships among native and non-native bumble bees (Bombus spp. Latreille) (Polidori and Nieves-Aldrey 2015), compare progeny size of Osmia bicornis L. (Seidelmann 2014) and Osmia lignaria Say (Helm et al. 2021) via provision studies, and explore flight morphology and metabolic rates of Megachile rotundata Fabricius (Grula et al. 2021). EPI is a composite variable that includes the ratio of mesosomal mass/whole body mass ( r ) and wing loading ( L ), which is a ratio of whole body mass/total wing area.

In this study, we calculated EPI for four native bee species from three different families (Andrenidae, Apidae, and Halictidae). Our objectives were to determine if EPI varied among sexes of solitary bees and whether these morphological measurements could be used to predict flight behavior. We hypothesized that EPI would differ between the sexes due to the variable ecological and social roles that male and female bees play.

## Materials and methods

## Bee collection

Agapostemon virescens Fabricius ( $\mathrm{N}=20 q$ and $20 \delta^{\top}$ ) and Andrena carlini Cockerell ( $\mathrm{N}=12+$ and $12 \delta^{\top}$ ) were collected by Sam Droege at the USGS Patuxent Wildlife Research Center. Melissodes bimaculata Lepeletier ( $\mathrm{N}=72+$ and $21 \circlearrowleft^{\top}$ ) were captured
near Scooba, Mississippi (Campbell et al. 2016). Xylocopa virginica L. ( $\mathrm{N}=19$ q and $18 \delta^{\lambda}$ ) were captured in St. Louis, Missouri in May 2015. All bees were either collected with sweep nets or colored pan traps. Additionally, all bees were collected from small locales to limit geographical/environmental differences in body size. These four species were chosen due to their commonness and easily distinguishable features.

## Specimen preparation and trait measurements

Whole specimens were dried for 72 hours at $45^{\circ} \mathrm{C}$ prior to determination of whole body mass (WBM) and mesosomal mass (MM) via a digital scale. One forewing and one hind wing were removed and flattened between glass microscope slides (Darveau 2005). Each mounted wing was photographed with a 5 mm size standard in the frame using a dissecting microscope (Zeiss). Images were assembled using Tps Utility software (Rohlf 2012), and wing area was determined with the TpsDIG software package (Rohlf 2005). We assumed symmetry between the paired wings, and therefore, we calculated total wing area (TWA) by doubling the forewing and hindwing areas that were measured from each specimen: TWA $=$ (forewing area*2) + (hindwing area*2).

## Excess power index calculation and statistical analysis

The excess power index (EPI) was calculated with the following formula:

$$
\text { EPI }=\sqrt{ } \frac{(\text { Mesosomal Mass } / \text { Whole Body Mass })^{2}}{\text { Whole Body Mass / Total Wing Area }}
$$

We performed pairwise comparisons of single and composite variables between males and females of each species using the Mann-Whitney $U$ test. To better understand the relative contributions of each portion of the EPI formula to differences between sexes, we compared two additional composite variables: 1. the ratio of mesosomal mass/whole body mass (r) and 2. the ratio of whole body mass/total wing area or wing loading (L). We judged statistical significance to be $\mathrm{p}<0.05$.

## Results

The whole body mass of females was significantly larger than males in all four bee species (Table 1). For $A$. virescens and $X$. virginica, the r and L values were both significantly different between females and males. Males had both an elevated r and lower L compared to females, both contributing positively to a male-biased dimorphism in EPI (Fig. 1, Table 1). In contrast, both M. bimaculata and A. carlini r values were not significantly different between male and female cohorts (Table 1). However, male L was significantly lower than females in both species, resulting in a male-biased dimorphism in EPI (Fig. 1, Table 1). Wing surface area was significantly different between sexes in all bee species except $X$. virginica. See Table 2 for all test statistics.

Table I. Mean morphological measurements ( $\pm$ SE) for four male and female native bee species that were used to calculate Excess Power Indices (EPI). An * indicates statistical significance between male and female of each bee species at $\mathrm{P}=0.05$.

| Bee Species/Sex | WBM (mg) | MM (mg) | r | WSA (mm ${ }^{\text {a }}$ | $\mathrm{L}\left(\mathrm{mg} / \mathrm{mm}^{2}\right)$ |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Agapostemon virescens $\mathrm{+}, \mathrm{~N}=20$ | 15.76 (0.76)* | 5.98 (0.28)* | 0.38 (0.005)* | 50.65 (1.06)* | 0.31 (0.009)* |
| Agapostemon virescens $\widehat{\top}^{\text {² }}, \mathrm{N}=20$ | 8.25 (0.36) | 3.61 (0.17) | 0.44 (0.006) | 36.20 (0.75) | 0.23 (0.007) |
| Andrena carlini +, $\mathrm{N}=12$ | 36.64 (3.03)* | 11.18 (.64)* | 0.31 (0.01) | 67.32 (1.60)* | 0.54 (0.04)* |
| Andrena carlini ${ }^{\text {O}}, \mathrm{N}=12$ | 11.02 (0.40) | 3.30 (0.30) | 0.30 (0.03) | 44.66 (1.47) | 0.25 (0.007) |
| Melissodes bimaculata ㅇ, $\mathrm{N}=72$ | 41.40 (0.90)* | 14.21 (0.30)* | 0.34 (0.003) | 87.14 (0.80)* | 0.47 (0.008)* |
| Melissodes bimaculata ${ }^{\text {® }}, \mathrm{N}=21$ | 27.66 (1.37) | 9.50 (0.44) | 0.35 (0.006) | 76.05 (1.53) | 0.36 (0.01) |
| Xylocopa virginica + , $\mathrm{N}=19$ | 259.17 (14.44)* | 68.96 (2.22) | 0.27 (0.01)* | 188.52 (3.72) | 1.36 (0.06)* |
| Xylocopa virginica ${ }^{\text {², }}, \mathrm{N}=18$ | 152.15 (4.97) | 62.29 (3.23) | 0.41 (0.02) | 196.69 (3.93) | 0.77 (0.02) |

$\mathrm{WBM}=$ whole body mass, $\mathrm{MM}=$ mesosomal mass, $\mathrm{r}=$ mesosomal/body mass ratio, WSA $=$ wing surface area, $\mathrm{L}=$ wing loading.


Figure I. Excess Power Indices (EPI) ( $\pm$ SE) of four native bee species (Agapostemon virescens, Andrena carlini, Melissodes bimaculata, Xylocopa virginica) and their corresponding sex. All Mann-Whitney U tests found that all males of each species had a significantly higher EPI compared to females at $\mathrm{p}<0.05$.

## Discussion

Differences in body size of male and female insects can result in different thermoregulatory and flight abilities (Gilchrist 1990). In general, total wing area in bees has been shown to be positively related to body mass (Bullock 1999) as might be expected. Body mass differences and wing loading in these four bee species were probably due to mating and foraging behaviors. Male bees invest no energy into their offspring; their activities are primarily directed to finding mates. Thus, male bee activity is driven by (1) the need to thermoregulate flight muscles, (2) finding enough nectar to power flight, and (3) find and mate with females (Willmer and Stone 2004). Thus, having a smaller body but increased wing loading should allow males to fly longer distances or remain in flight for longer time periods in search of females but also limit the amount of nectar

Table 2. Mann-Whitney U test statistics for morphological parameters and Excess Power Indices.

|  | WBM | MM | $\mathbf{r}$ | WSA | $\mathbf{L}$ | EPI |
| :--- | :---: | :---: | :---: | :---: | :---: | :---: |
| Andrena carlini | $z=4.13$, | $z=4.13$, | $z=0.14$, | $z=4.13$, | $z=4.13$, | $z=2.45$, |
|  | $\mathrm{P}<0.00001$ | $\mathrm{P}<0.00001$ | $\mathrm{P}=0.89$ | $\mathrm{P}<0.00001$ | $\mathrm{P}<0.00001$ | $\mathrm{P}=0.014$ |
| Melissodes bimaculata | $z=5.87$, | $z=5.93$, | $z=0.11$, | $z=5.22$, | $z=5.80$, | $z=3.92$, |
|  | $\mathrm{P}<0.00001$ | $\mathrm{P}<0.00001$ | $\mathrm{P}=0.91$ | $\mathrm{P}<0.00001$ | $\mathrm{P}<0.00001$ | $\mathrm{P}=0.001$ |
| Agapostemon virescens | $z=5.26$, | $z=4.93$, | $z=4.65$, | $z=5.37$, | $z=4.75$, | $z=5.32$, |
|  | $\mathrm{P}<0.00001$ | $\mathrm{P}<0.00001$ | $\mathrm{P}<0.00001$ | $\mathrm{P}<0.00001$ | $\mathrm{P}<0.00001$ | $\mathrm{P}<0.00001$ |
| Xylocopa virginica | $z=4.85$, | $z=1.84, \mathrm{P}=$ | $z=4.60$, | $z=1.14$, | $z=5.18$, | $z=4.60$, |
|  | $\mathrm{P}<0.00001$ | 0.066 | $\mathrm{P}<0.00001$ | $\mathrm{P}=0.25$ | $\mathrm{P}<0.00001$ | $\mathrm{P}<0.00001$ |

needed to sustain themselves. Additionally, many male bee species (e.g., A. carlini) will 'patrol' flowers in search of females (Schrader and LaBerge 1978). Male bees, including A. virescens and M. bimaculata will sleep inside flowers (Rau 1938; Abrams and Eickwort 1981). This male-biased dimorphism in EPI, a composite variable that describes the predicted flight capacity of an individual based on mass and morphological attributes, can be used to predict flight behavior between bee sexes.

Female bee activities are primarily driven by the need to collect floral resources (e.g., pollen and nectar) to provision themselves and for their offspring. Unlike social bee species, solitary females complete all nest construction and provisioning on their own. Although some species in this study are considered communal nesters, each individual female only provisions her own nest(s). Solitary bees provision each brood cell sequentially and must provide enough food resources for each one (mass provisioning), which will take multiple foraging trips per brood cell (Danforth et al. 2019). Having a larger body mass may enable females to better thermoregulate body temperature enabling them to forage in less than optimal conditions (Stone 1993). Additionally, a larger body size should also allow for carrying larger loads of pollen and nectar and the ability to fly to farther flower patches (Skandalis et al. 2009). Indeed, larger-bodied female bee species have been correlated with increased foraging ranges (Greenleaf et al. 2007). However, female bees may be faced with an evolutionary tradeoff; larger body sizes can carry more provisions but have reduced flight capacity (Seidelmann 2014). Alternatively, larger body size in female bees has not always shown consistent patterns when relating body size to fecundity (Tepedino et al. 1984) and may contribute to higher parasitism rates by parasitoids (Müller et al. 1996). Thus, multiple evolutionary pressures may contribute to bee body size.

Among the bee species utilized for this study, $X$. virginica, a cavity nesting species, is probably the most thoroughly studied. Males often compete in intrasexual aggression which is mediated primarily through the body size of the males (Barthell and Baird 2004). The interactions between these males are often done in the form of aerial pursuits, which indicates the necessity for the males to have a better EPI in order to fly more efficiently for these male aggression encounters (Barthell and Baird 2004). Interestingly, despite the difference in EPI between male and female $X$. virginica, wing area did not significantly differ. The ecologies of the other three bee species in this study are
poorly documented. However, they are all ground nesters but include a range of nesting habits from communal nesters for $A$. virescens (Eickwort 1981) to solitary nests for A. carlini (Schrader and LaBerge 1978). All four bee species feed on a variety of plants for pollen and nectar and, thus, are considered polylectic (Mitchell 1960, 1962).

## Conclusion

The four bee species utilized for this study comprised three ground-nesting species (A. carlini, M. bimaculata, A. virescens) and one cavity nesting bee (X. virginica). Despite the differences in nesting and the wide taxonomic differences, the morphological measurements between the sexes showed similar trends. Although we only assessed four species, this proof of concept would also be expected for other solitary bees that have similar life histories and ecologies. To our knowledge, this is the first report of EPI measurements on non-Apis/Bombus bees other than O. bicornis, O. lignaria, and M. rotundata (Seidelmann 201; Grula et al. 20214; Helm et al. 2021) and the only wild bee species utilized from the Nearctic for this type of study. Utilizing EPI for other wild bee species may allow for inferences to be made regarding male/female behaviors but also comparative studies across landscapes and environmental conditions.

## Acknowledgements

We thank Sam Droege of the USGS Patuxent Wildlife Research Center for providing specimens. This work was completed at High Point University.

## References

Abbate A, Campbell JW, Kimmel CB, Kern Jr WH (2019) Urban development decreases bee abundance and diversity within coastal dune systems. Global Ecology and Conservation 20: 1-13. https://doi.org/10.1016/j.gecco.2019.e00711
Abrams J, Eickwort GC (1981) Nest switching and guarding by the communal sweat bee Agapostemon virescens (Hymenoptera, Halictidae). Insectes Sociaux 28: 105-116. https://doi.org/10.1007/BF02223699
Allen-Perkins A, Magrach A, Dainese M, Garibaldi LA, Kleijn D, Rader R, Reilly JR, Winfree R, Lundin O, McGrady CM, Brittain C, Biddinger D, Artz DR, Elle E, Hoffman G, Ellis JD, Daniels J, Gibbs J, Campbell JW, et al. (2022) CropPol: a dynamic, open and global database on crop pollination. Ecology 103(3): e3614. https://doi.org/10.1002/ecy. 3614
Barthell JF, Baird TA (2004) Size variation and aggression among male Xylocopa virginica (L.) (Hymenoptera: Apidae) at a nesting site in central Oklahoma. Journal of the Kansas Entomological Society 77: 10-20. https://doi.org/10.2317/0301-27.1
Bullock SH (1999) Relationships among body size, wing size and mass in bees from a tropical dry forest in Mexico. Journal of the Kansas Entomological Society 72: 426-439.

Cameron SA, Lozier JD, Strange JP, Koch JB, Cordes N, Solter LF, Griswold TL (2011) Patterns of widespread decline in North American bumble bees. Proceedings of the National Academy of Sciences 108: 662-667. https://doi.org/10.1073/pnas. 1014743108
Campbell JW, Miller DA, Martin JA (2016) Switchgrass (Panicum virgatum) intercropping within managed loblolly pine (Pinus taeda) does not affect wild bee communities. Insects 7(4): 62. https://doi.org/10.3390/insects7040062
Colla SR, Gadallah F, Richardson L, Wagner D, Gall L (2012) Assessing declines of North American bumble bees (Bombus spp.) using museum specimens. Biodiversity and Conservation 21: 3585-3595. https://doi.org/10.1007/s10531-012-0383-2
Danforth BN, Minckley RL, NeffJL, Fawcett F (2019) The Solitary Bees: Biology, Evolution, Conservation. Princeton University Press, 472 pp. https://doi.org/10.1515/9780691189321
Darveau CA, Hochachka PW, Welch Jr KC, Roubik DW, Suarez RK (2005) Allometric scaling of flight energetics in Panamanian orchid bees: a comparative phylogenetic approach. Journal of Experimental Biology 208: 3581-3591. https://doi.org/10.1242/jeb. 01776
Eickwort GC (1981) Aspects of the nesting biology of five Nearctic species of Agapostemon (Hymenoptera: Halictidae). Journal of the Kansas Entomological Society 54: 337-351.
Gilchrist GW (1990) The consequences of sexual dimorphism in body size for butterfly flight and thermoregulation. Functional Ecology 4: 475-487. https://doi.org/10.2307/2389315
Greenleaf SS, Williams NM, Winfree R, Kremen C (2007) Bee foraging ranges and their relationship to body size. Oecologia 153: 589-596. https://doi.org/10.1007/s00442-007-0752-9
Grula CC, Rinehart JP, Greenlee KJ, Bowsher JH (2021) Body size allometry impacts flightrelated morphology and metabolic rates in the solitary bee Megachile rotundata. Journal of Insect Physiology 133: 104275. https://doi.org/10.1016/j.jinsphys.2021.104275
Helm BR, Baldwin MA, Rinehart JP, Yocum GD, Greenlee KJ, Bowsher JH (2021) Body and wing allometries reveal flight-fecundity tradeoff in response to larval provisioning in Osmia lignaria (Hymenoptera: Megachilidae). Journal of Insect Science 21(3): 11. https://doi. org/10.1093/jisesa/ieab035
Hepburn HR, Radloff SE, Steele GR, Brown RE (1998) Dimensional aspects of flight in the honey bees of Africa. Journal of Apicultural Research 37: 147-154. https://doi.org/10.10 80/00218839.1998.11100966
Hepburn HR, Radloff, SE, Fuchs S (1999) Flight machinery dimensions of honeybees, Apis mellifera. Journal of Comparative Physiology B 169: 107-112. https://doi.org/10.1007/ s003600050200
Kleijn D, Raemakers I (2008) A retrospective analysis of pollen host plant use by stable and declining bumble bee species. Ecology 89: 1811-1823. https://doi.org/10.1890/07-1275.1
Michener C (2007) The Bees of the World. $2^{\text {nd }}$ edn. John Hopkins University Press, Baltimore, 972 pp. https://doi.org/10.56021/9780801885730
Mitchell TB (1960) Bees of the Eastern United States. North Carolina Agricultural Experiment Station Technical Bulletin No. 141.
Mitchell TB (1962) Bees of the Eastern United States. North Carolina Agricultural Experiment Station Technical Bulletin No. 152.
Müller CB, Blackburn TM, Schmid-Hempel P (1996) Field evidence that host selection by conopid parasitoids is related to host body size. Insectes Sociaux 43: 227-233. https://doi. org/10.1007/BF01242924

Ndungu NN, Kiatoko N, Ciosi M, Salifu D, Nyansera D, Masiga D, Raina SK (2017) Identification of stingless bees (Hymenoptera: Apidae) in Kenya using morphometrics and DNA barcoding. Journal of Apicultural Research 56: 341-353. https://doi.org/10.1080/00218 839.2017.1327939

Ollerton J, Winfree R, Tarrant S (2011) How many flowering plants are pollinated by animals? Oikos 120: 321-326. https://doi.org/10.1111/j.1600-0706.2010.18644.x
Polidori C, Nieves-Aldrey JL (2015) Comparative flight morphology in queens of invasive and native Patagonian bumblebees (Hymenoptera: Bombus). Comptes Rendus Biologies 338: 126-133. https://doi.org/10.1016/j.crvi.2014.11.001
Radloff SE, Hepburn HR, Koeniger G (2003) Comparison of flight design of Asian honeybee drones. Apidologie 34: 353-358. https://doi.org/10.1051/apido:2003031
Rau P (1938) Additional observations on the sleep of insects. Annals of the Entomological Society of America 31: 540-556. https://doi.org/10.1093/aesa/31.4.540
Rohlf FJ (2012) Tps Utility Program, ver. 1.50. Department of Ecology and Evolution, Stony Brook, State University of New York, New York.
Rohlf FJ (2005) TpsDig, digitize landmarks and outlines, ver. 2.04. Department of Ecology and Evolution, Stony Brook, State University of New York, New York.
Scheper J, Reemer M, van Kats R, Ozinga WA, van der Linden GT, Schaminée JH, Siepel H, Kleijn D (2014) Museum specimens reveal loss of pollen host plants as key factor driving wild bee decline in The Netherlands. Proceedings of the National Academy of Sciences 111: 17552-17557. https://doi.org/10.1073/pnas. 1412973111
Schrader MN, LaBerge WE (1978) The nest biology of the bees Andrena (Melandrena) regularis Malloch and Andrena (Melandrena) carlini Cockerell (Hymenoptera: Andrenidae). Biological Notes No. 108. Illinois Natural History Survey, 24 pp. https://doi.org/10.5962/ bhl.title. 15182
Skandalis DA, Tattersall GJ, Prager S, Richards MH (2009) Body size and shape of the large carpenter bee, Xylocopa virginica (L.) (Hymenoptera: Apidae). Journal of the Kansas Entomological Society 82: 30-42. https://doi.org/10.2317/JKES711.05.1
Seidelmann K (2014) Optimal progeny body size in a solitary bee, Osmia bicornis (Apoidea: Megachilidae). Ecological Entomology 39: 656-663. https://doi.org/10.1111/een. 12145
Stone GN (1993) Thermoregulation in four species of tropical solitary bees: the roles of size, sex and altitude. Journal of Comparative Physiology B 163: 317-326. https://doi. org/10.1007/BF00347782
Tepedino VJ, Thompson R, Torchio PF (1984) Heritability for size in the megachilid bee Osmia lignaria propinqua Cresson. Apidologie 15: 83-88. https://doi.org/10.1051/apido:19840108
Vaudo AD, Fritz ML, López-Uribe MM (2018) Opening the door to the past: Accessing phylogenetic, pathogen, and population data from museum curated bees. Insect Systematics and Diversity 2(5): 4. [1-14] https://doi.org/10.1093/isd/ixy014
Willmer PG, Stone GN (2004) Behavioral, ecological, and physiological determinants of the activity patterns of bees. Advances in the Study of Behavior 34: 347-466. https://doi. org/10.1016/S0065-3454(04)34009-X


[^0]:    * These authors contributed equally.

